

Differentiation induction of murine erythroleukemia cells by butylated hydroxytoluene

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Received 15 November 1983

Butylated hydroxytoluene (BHT) which is widely used as an anti-oxidant in food has been found to induce the differentiation of murine erythroleukemia cells. BHT also amplifies the differentiation inducing activity of DMSO.

Butylated hydroxytoluene Differentiation Erythroleukemia cell DMSO

1. INTRODUCTION

During the studies on the differentiation-inducing substances of animal cells, we found that butylated hydroxytoluene (BHT) induces the differentiation of murine erythroleukemia cells.

BHT is widely used as an antioxidant or a stabilizer in food, feed, cosmetics and pharmaceutical products. Recently it has been reported that BHT shows a variety of biological activities; e.g., inactivation of viruses [2,3], antimutagenicity [4], anti-carcinogenesis [5-7], and stimulation of DNA synthesis [8].

We report here that BHT also induces the differentiation of murine erythroleukemia cells with high efficiency, and amplifies the differentiation-inducing activity of DMSO.

2. MATERIALS AND METHODS

2.1. Materials

Butylated hydroxytoluene (BHT) was purchased from Wako Chemical Co. (Tokyo). Ham's F12

Studies on the differentiation inducing substances of animal cells II. See [1]

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medium and fetal bovine serum were obtained from GIBCO and Flow Laboratory, respectively. DMSO was a product of Junsei Chemical Co. (Tokyo).

2.2. Cell line and cell culture

Mouse erythroleukemia cells (clone B8) [9] were kindly provided by Dr K. Nose. Cells were grown in Ham's F12 medium supplemented with 15% fetal bovine serum and kanamycin (60 µg/ml) at 37°C in a humidified incubator with 8% CO₂ atmosphere.

2.3. Differentiation assay

The cells (6×10^4 cells/ml) were cultured for 3-7 days at 37°C with the indicated concentrations of BHT (see text) and/or 1.5% DMSO. The differentiated cells were determined by the modified benzidine-staining method [10].

3. RESULTS AND DISCUSSION

3.1. Effect of BHT on growth and differentiation of mouse erythroleukemia cells

After the erythroleukemia cells were cultured with the various concentrations of BHT for 3 days, the differentiated cells were determined by the benzidine-staining method. Fig.1 shows that the

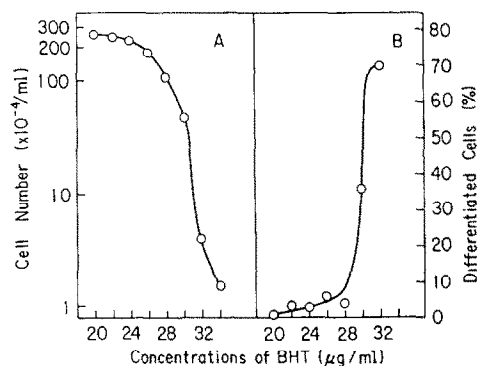


Fig. 1. Effect of BHT on growth (A) and differentiation (B) of mouse erythroleukemia (B8) cells. The cells (6×10^4 cells/ml) were cultured for 3 days at 37°C with the indicated concentrations of BHT in 5 ml of Ham's F12 medium supplemented with 15% fetal bovine serum in humidified air containing 8% CO_2 . The differentiated cells were examined by the benzidine-staining method [9].

differentiation occurs at about $30 \mu\text{g/ml}$ of BHT maximally.

3.2. Time-course analysis of the differentiation induced by BHT and DMSO

Time-course of differentiation was examined (fig. 2). Maximum cell differentiation by BHT was observed after 3 days culture, whereas maximum differentiation induction by DMSO was observed after 6 days culture.

3.3. Cooperative effect of BHT and DMSO on the differentiation of erythroleukemia cells

Fig. 3 shows the cooperative effect of 1.5%

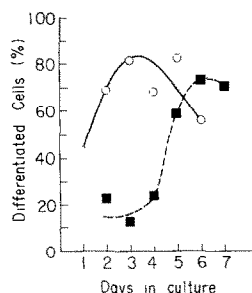


Fig. 2. Time-course analysis of differentiation by $30 \mu\text{g/ml}$ of BHT (○) and 1.5% DMSO (■). The culture conditions and examination method of the differentiated cells were the same as described in the legend of fig. 1, except the days in culture.

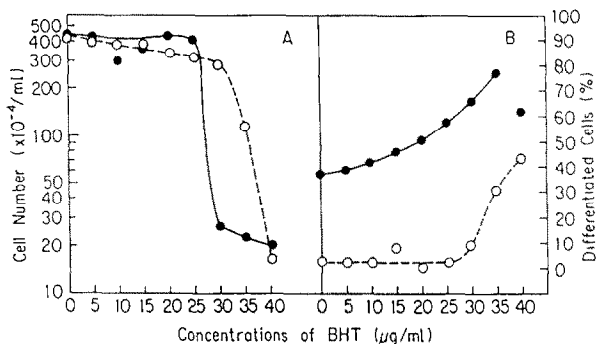


Fig. 3. Cooperative effect of 1.5% DMSO and the different concentrations of BHT on growth (A) and differentiation (B) of mouse erythroleukemia (B8) cells. The cells were cultured for 6 days with 1.5% DMSO and the indicated concentrations of BHT (●), and with BHT only (○). The experimental details are described in the legend of fig. 1, except the days in culture (this experiment was repeated 3 times, and the results were confirmed).

DMSO and the indicated concentrations of BHT on the growth (A) and the differentiation (B) of mouse erythroleukemia (B8) cells. The cells were cultured for 6 days with 1.5% DMSO and the various concentrations of BHT (●), and with BHT only (○). This figure shows inducibility of about 0% with BHT at the concentration range of 5–25 $\mu\text{g/ml}$, and of about 40% with DMSO, respectively. Inducibilities, however, by the cooperation of DMSO and BHT (5–25 $\mu\text{g/ml}$) were higher than the sums of inducibilities caused by the individual inducers. Somewhat weak synergism was observed.

BHT is a strong antioxidant. Other antioxidants such as retinoic acid [11–13], tocopherol [7,14] and ascorbic acid [7,15] are also known to inhibit carcinogenesis or to influence cell differentiation. The redox system might take part in the mechanisms of differentiation and carcinogenesis.

BHT has been reported to possess a variety of biological activities; e.g., inactivation of viruses [2,3], anti-mutagenic activity [4], inhibition of physical [5] and chemical [6,7] carcinogenesis and the stimulation of DNA synthesis [8]. It was recently reported that BHT inhibits cell differentiation of mouse myeloid leukemia (M1) cells [16] induced by dexamethasone at the concentration of $10 \mu\text{g/ml}$ [17].

We have described here that BHT is a highly ef-

ficient inducer of differentiation. The fact that BHT, a widely used reagent, has differentiation controlling activity might present a serious problem to embryogenesis or development of animals.

REFERENCES

- [1] Asahi, K., Ono, I., Kusakabe, H., Nakamura, G. and Isono, K. (1981) *J. Antibiotics* 34, 919-920.
- [2] Snipes, W., Person, S., Keith, A. and Cupp, J. (1975) *Science* 188, 64-66.
- [3] Brugh, M., jr (1977) *Science* 197, 1291-1292.
- [4] Calle, L.M., Sullivan, P.D., Nettleman, M.D., Ocasio, I.J., Blazyk, J. and Jollick, J. (1978) *Biochem. Biophys. Res. Commun.* 85, 351-356.
- [5] Black, H.S., Chan, J.T. and Brown, G.E. (1978) *Cancer Res.* 38, 1384-1387.
- [6] McCay, P.B., King, M.M. and Pitha, J.V. (1981) *Cancer Res.* 41, 3745-3748.
- [7] Slaga, T.J. and Bracken, W.M. (1977) *Cancer Res.* 37, 1631-1635.
- [8] Larsen, J.C. and Tarding, F. (1978) *Toxicological Aspects of Food Safety Arch. Toxicol.*, suppl.1, 147-150.
- [9] Ostertag, W., Crozier, T., Kuge, N., Melderis, H. and Dube, S. (1973) *Nat. New Biol.* 243, 203-205.
- [10] Orkin, S., Haroshi, F.I. and Leder, P. (1975) *Proc. Natl. Acad. Sci. USA* 71, 98-102.
- [11] Longnecker, D.S., Curphey, T.J., Kuhlmann, E.T. and Roebuck, B.D. (1982) *Cancer Res.* 42, 19-24.
- [12] Strickland, S. and Mahdavi, V. (1978) *Cell* 15, 393-403.
- [13] Takenaga, K., Hozumi, M. and Sakagami, Y. (1980) *Cancer Res.* 40, 914-919.
- [14] Prasad, K.N., Ramanujam, S. and Gaudreau, D. (1979) *Proc. Soc. Exp. Biol. Med.* 161, 570-573.
- [15] Eguchi, G., Masuda, A., Karasawa, Y., Kodama, R. and Itoh, Y. (1982) in: *Stability and Switching in Cellular Differentiation* (Clayton, R.M. and Truman, D.E.S. eds) pp. 209-221, Plenum, New York.
- [16] Ichikawa, Y. (1969) *J. Cell. Physiol.* 74, 223-234.
- [17] Takenaga, K., Honma, Y. and Hozumi, M. (1981) *Gann* 72, 104-112.